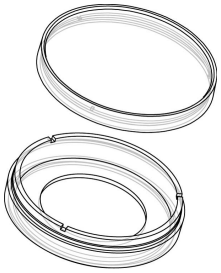


The ibidi product family is comprised of a variety of µ–Slides and µ–Dishes, which have all been designed for high–end microscopic analysis of fixed or living cells. The high optical quality of the material is similar to that of glass, so you can perform all kinds of fluorescence experiments with uncompromised resolution and choice of wavelength.

The µ–Dish 35mm, low allows you to perform high resolution microscopy in a 35 mm Petri–dish with 7 mm walls. The low height makes high numerical apertures of Köhler illumination possible and provides large access for micromanipulation. The lid can be closed to hinder evaporation during long term experiments.

Material

ibidi µ–Slides, µ–Dishes, and µ–Plates are made of a plastic that has the highest optical quality. The bottom material exhibits extremely low birefringence and autofluorescence, similar to that of glass. Also, it is not possible to detach the bottom from the upper part. The µ–Slides, µ–Dishes, and µ–Plates are not autoclavable, since they are only temperature–stable up to 80°C/175°F. Please note that gas exchange between the medium and incubator's atmosphere occurs partially through the polymer coverslip, which should not be covered.

Optical Properties ibidi Standard Bottom	
Refractive index n_D (589 nm)	1.52
Abbe number	56
Thickness	No. 1.5 (180 µm)
Material	microscopy plastic/ polymer coverslip

Please note! The ibidi Standard Bottom is compatible with certain types of immersion oil only. A list of suitable oils can be found on page 3.

Geometry

Geometry of the µ–Dish 35mm, low	
Diameter dish	35 mm
Volume	800 µl
Growth area	3.5 cm^2
Diameter growth area	21 mm
Coating area using 400 µl	4.2 cm^2
Height with / without lid	9 mm / 7 mm
Bottom matches coverslip	No. 1.5

Shipping and Storage

The µ–Slides, µ–Dishes and µ–Plates are sterilized and welded in a gas-permeable packaging. The shelf life under proper storage conditions (in a dry place, no direct sunlight) is listed in the following table.

Conditions	
Shipping conditions	Ambient
Storage conditions	RT (15-25°C)

Shelf Life of Different Surfaces	
ibiTreat, Glass Bottom, ESS	36 months
Collagen, Poly-Lysine	18 months
Fibronectin	4 months

Surface and Coating

The µ–Dish 35mm, low is available with ibiTreat and uncoated surface. The ibiTreat surface is a physical treatment and optimized for adhesion of most cell types. Many cell lines as well as primary cells were tested for good cell growth. The uncoated surface is a very hydrophobic surface and allows no direct cell growth. It is suitable for specific coatings or suspension cells.

If you like to establish a particular coating for your demands we recommend to test your coating procedure on uncoated and ibiTreat µ–Dishes, since some biomolecules adhere differently to hydrophobic or hydrophilic plastic surfaces.

- Prepare your coating solution according to the manufacturer's specifications or reference. Prepare your µ–Dish, ibiTreat or uncoated. Adjust the concentration to a coating area of 4.2 cm^2 and 400 µl.
- Apply 400 µl into the growth area. Make sure that the entire bottom is covered with liquid easily tilting or shaking the µ–Dish. Put on the lid and leave at room temperature for at least 30 minutes.

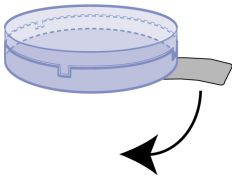
- Aspirate the solution and wash. Optionally, let dry at room temperature.

Detailed information about coatings is provided in Application Note 08 "Cell culture coating".

The μ–Dish 35mm, high is also available with an elastically supported surface (ESS) and a glass bottom surface. Please refer to the instructions for detailed information.

Protection Film

Remove the protection film before usage!

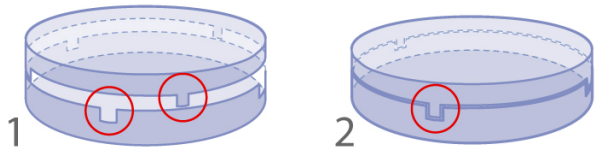


The bottom of the μ–Dish is covered with a film to protect the optical quality of the plastic surface. Please pull off the protection film before usage!

The protection film of all μ–Dishes is color-coded:

Color	Surface	Variation
Blue	ibiTreat	plain
Red	ibiTreat	with Grid–500
Yellow	ibiTreat	with Culture-Insert
White	uncoated	plain
Black	uncoated	with Grid–500
Green	uncoated	with Culture–Insert

Using The Lid



1. open position, easy opening
2. close position, for long term studies, minimal evaporation

Seeding Cells

Depending on your cell type, application of a 4–9 × 10^4 cells/ml suspension should result in a confluent layer within 2–3 days.

- Trypsinize and count cells as usual. Dilute the cell suspension to the desired concentration.
- Apply 400 µl cell suspension into the inner well of the μ–Dish. Avoid shaking as this will result in inhomogeneous distribution of the cells.
- After cell attachment add additionally 400 µl of pure medium to ensure optimal grow conditions.
- Cover the μ–Dish with the supplied lid. Incubate at 37°C and 5 % CO_2 as usual.

We recommend not to fill more than the indicated total volume into the μ–Dish 35mm, low in order to avoid the liquid contacting the lid.

Undemanding cells can be left in their seeding medium for several days and grow to confluence there. However, best results are achieved when the medium is changed every 2–3 days. Carefully aspirate the old medium and replace it by up to 800 µl fresh medium.

Tip:

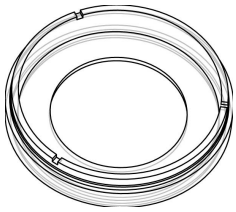
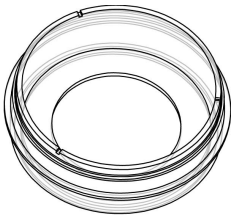
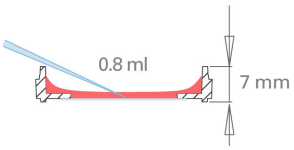
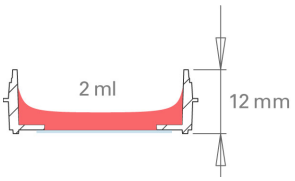
You can stack the μ–Dishes to save space in your incubator. This will not affect cell growth. We recommend making batches with up to 6 μ–Dishes, due to stability reasons. Placing the μ–Dishes into larger Petri dishes simplifies transport and prevents evaporation, heat loss, and contamination when the incubator is opened.

Preparation for Cell Microscopy

To analyze your cells no special preparations are necessary. Cells can be observed live or fixed directly in the μ–Dish preferably on an inverted microscope. You can use any fixative of your choice. The μ–Dish material is compatible with a variety of chemicals, e.g. Acetone or Methanol. Further specifications can be found at www.ibidi.com. Due to the thin bottom high resolution microscopy is possible.

For optimal results in fluorescence microscopy and storage of stained probes, ibidi provides a mounting medium optimized for μ–Dishes and μ–Slides (ibidi Mounting Medium, 50001).

µ–Dish 35mm Selection Guide

µ–Dish 35mm, low	µ–Dish 35mm, high
Low walls (7 mm) for large access to the cells. Designed for micromanipulation and microinjection.	High walls (12 mm) for all standard applications. Also available with glass bottom, relocation grid, and elastic surface (ESS).
	
	

Minimizing Evaporation

Using the µ–Dish with a closed lid, the evaporation in an incubator system with 37°C and 95 % humidity is around 1 % per day. Using the µ–Dish with a closed lid in a 37°C heating system with low humidity (between 20 % and 40 %), the evaporation is around 10 % per day. For reducing the evaporation down to 1 % per day in all systems, we recommend sealing the lid with ibidi Anti–Evaporation Oil (50051).

Immersion Oil

When using oil immersion objectives, use only the immersion oils specified in the table. The use of a non–recommended oil could lead to the damage of the plastic material and the objective.

Company	Product	Ordering Number
Zeiss	Immersol 518 F	(Zeiss) 444960
Zeiss	Immersol W 2010	(Zeiss) 444969
Leica	Immersion liquid	(Leica) 11513859

Selected References

L. P. Frenzel, Z. Abdullah, A. K. Kriegeskorte, R. Dieterich, N. Lange, D. H. Busch, M. Kronke, O. Utermohlen, J. Hescheler, and T. Saric. Role of NKG2D–ligands and ICAM–1 in NK cell-mediated Lysis of Murine Embryonic Stem Cells and Embryonic Stem Cell–derived Cardiomyocytes. *Stem Cells*, 2008. doi: 10.1634/stemcells.2008-0528.

T. Quast, F. Eppler, V. Semmling, C. Schild, Y. Homsi, S. Levy, T. Lang, C. Kurts, and W. Kolanus. CD81 is Essential for the Formation of Membrane Protrusions and Regulates Rac1-Activation in Adhesion-Dependent Immune Cell Migration. *Blood*, 2011. doi: 10.1182/blood-2010-12-326595.

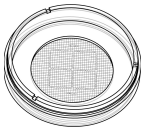
K. Rönnebäumer, U. Groß, and W. Bohne. The nascent parasitophorous vacuole membrane of *E. cuniculi* is formed by host cell lipids and contains pores which allow nutrient uptake. *Eukaryotic Cell*, 2008. doi: 10.1128/ec.00004-08.

µ–Dish 35mm, low Family

µ–Dish 35mm, low

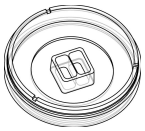
Cat. No.	Description	Characteristics
80136	**µ–Dish 35mm, low ibiTreat**: ⌀ 35 mm, low wall (800 µl volume), #1.5 polymer coverslip, tissue culture treated	hydrophilic, sterilized
80131	**µ–Dish 35mm, low Uncoated**: ⌀ 35 mm, low wall (800 µl volume), #1.5 polymer coverslip	hydrophobic, sterilized

µ–Dish 35mm, low Grid-500



Cat. No.	Description	Characteristics
80156	**µ–Dish 35mm, low ibiTreat Grid-500**: ⌀ 35 mm, high wall (800 µl volume), #1.5 polymer coverslip, tissue culture treated, grid repeat distance 500 µm	hydrophilic, sterilized
80151	**µ–Dish 35mm, low Uncoated Grid-500**: ⌀ 35 mm, high wall (800 µl volume), #1.5 polymer coverslip, grid repeat distance 500 µm	hydrophobic, sterilized

Culture Insert in µ–Dish 35mm, low **



Cat. No.	Description	Characteristics
80206	**Culture Insert in µ–Dish 35mm, low, ibiTreat**: ready to use, tissue culture treated	hydrophilic, sterilized
80201	**Culture Insert in µ–Dish 35mm, low, uncoated**: ready to use, tissue culture treated	hydrophobic, sterilized

** Please also refer to the Culture-Insert instructions.

For research use only!

Further technical specifications can be found at www.ibidi.com. For questions and suggestions please contact us by e-mail *info@ibidi.de* or by telephone +49 (0)89/520 4617 0. All products are developed and produced in Germany.