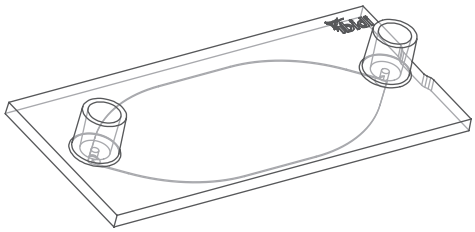
sticky-Slide Tissue

Instruction Manual



The sticky-Slide Tissue supports immunofluorescence and immunohistochemistry staining protocols of tissue sections on microscope glass slides.

This document applies to the following product:

80518	**sticky-Slide Tissue**

Material

The material of sticky-Slides is identical to that of µ-Slides. All sticky-Slides are delivered sterilized and individually packed. Please keep in mind that sterility is lost when non-sterile substrates are used. The sticky-Slides are not autoclavable, as they are only temperature-stable up to 60 ℃/140 ℉.
The sticky bottom itself is a 85 µm biocompatible double-faced adhesive tape. The tape is covered by a protection film, which must be removed before usage.

Shipping and Storage

The sticky-Slides are sterilized and sealed in a gas-permeable packaging. The shelf life under proper storage conditions (in a dry place, no direct sunlight) is outlined in the following table.

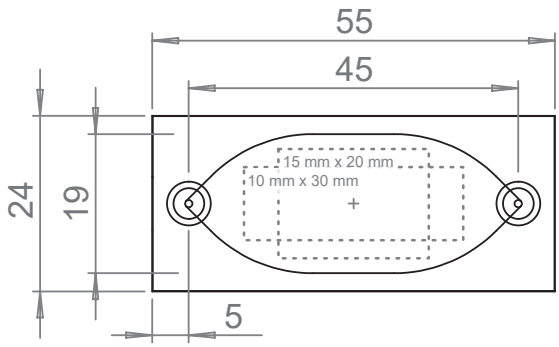
Conditions	
Shipping conditions	Ambient
Storage conditions	RT (15–25 ℃)

Shelf Life	
sticky-Slides	36 months

Geometry

Please note that the outer dimensions of the sticky-Slide Tissue are smaller compared to the standard ISO 8037/1 slide format.

Specifications	
Outer dimensions	24.0 × 55.0 mm^2
Total height	7.8 mm
Channel height	235 µm
Channel volume	160 µl
Ceiling thickness	1.67 mm
Adapters	Female Luer
Bottom	None



The sticky-Slide Tissue is made for tissue sections smaller than approx. 10 mm x 30 mm or 15 mm x 20 mm, respectively.

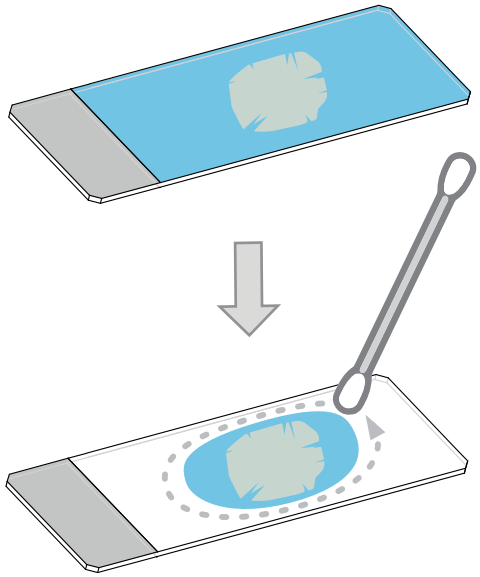
Surface Compatibility

The sticky-Slide Tissue is compatible with flat, clean, dust-free, fat-free surfaces, such as microscope slides and coverslips. Best results are achieved with completely dry surfaces. Please test your specific surface with a free sample from ibidi.com.

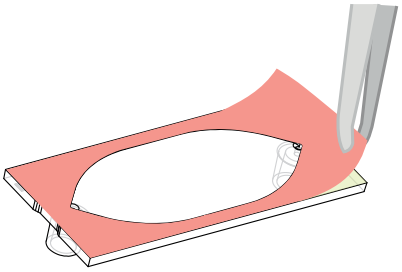
Handling and Assembly

Follow the protocol below to mount the sticky-Slide Tissue to a microscope slide bearing a tissue section after deparaffinization and rehydration.

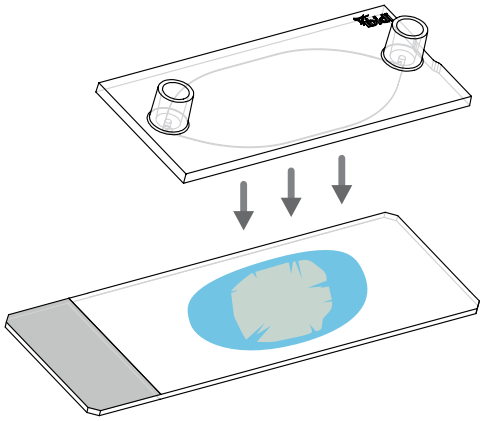
1. Deparaffinize and rehydrate your tissue.
2. Make sure the size of the tissue section is smaller than approx. 10 mm × 30 mm or 15 mm × 20 mm, respectively.
3. Clean and dry the surface around the tissue section. Do not touch the tissue section in order to keep it flat and even.



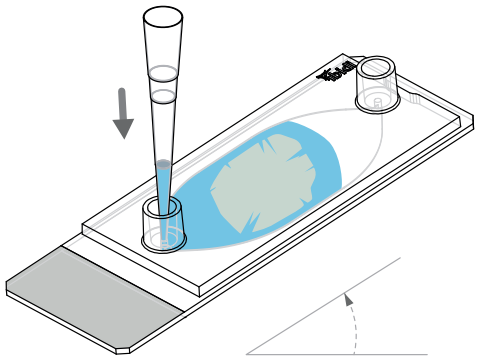
4. Remove the protection film from the sticky-Slide. First, scrape off a corner of the protection film using a scalpel or pointy tweezers. After that, grab this corner and remove the protection film.



5. Mount microscope slide and sticky-Slide by pressing firmly with your fingers (use gloves) until both parts are completely sealed. Make sure the tissue section is aligned with the channel. For a better visualization of the tissue section, a dark background is beneficial.



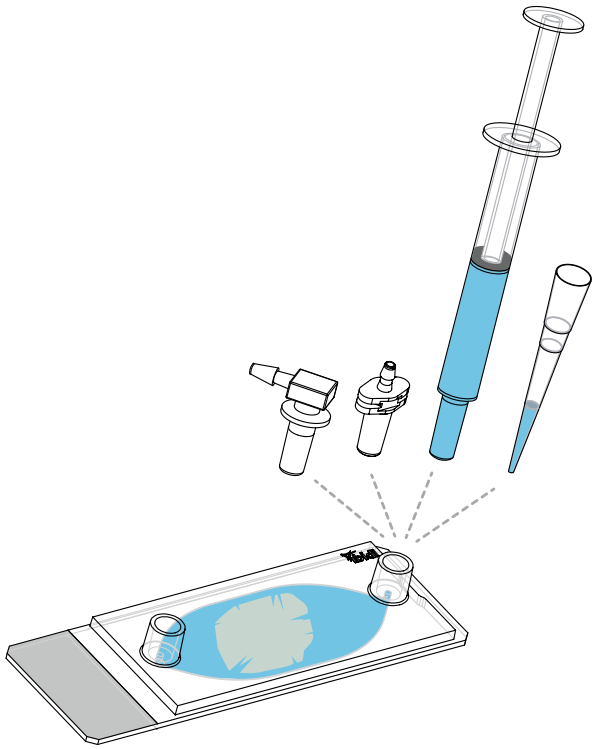
6. Fill the channel using a syringe or a pipet. Incline the slide and inject the liquid in an upwards motion so that air can escape above the liquid level. Optionally, connect Luer adapters and tubing for automated liquid exchange.



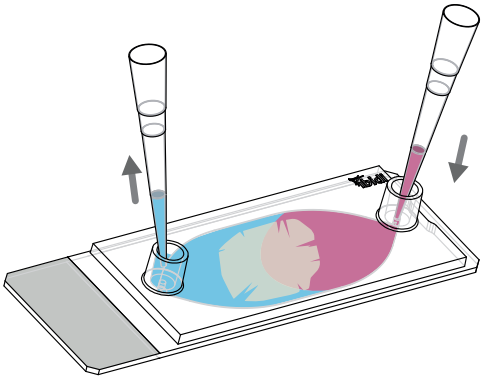
TIP – When filling the channel with a pipet, point the pipet tip directly onto the channel opening and inject the liquid.

TIP – Inclining the slide helps filling the channel without entrapped air bubbles. Keep in mind that air is much lighter than water. Let the air escape above the liquid.

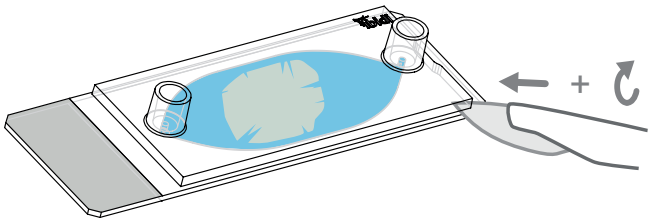
TIP – In case air bubbles become a problem during filling, use the final re-hydration solution (typically approx. 40–70% ethanol) to fill the channel. Subsequently, wash with water or buffer to remove the ethanol.



- For liquid exchange, a continuous exchange is recommended without removing any liquid from the channel itself: Add new liquid from one side removing it from the other. For a thorough wash, flush the channel with approx. 3× the channel volume, i.e. 480 µl.



- Follow your staining protocol.
- Optionally, remove the sticky-Slide Tissue with a scalpel and continue with mounting for microscopy.



TIP – Instead of using a microscope glass slide, a coverslip may be used to support high resolution microscopy.

Chemical Compatibility

The following table provides basic information on the chemical and solvent compatibility of the sticky-Slide Tissue. For a full list of compatible solvents and more information on chemical compatibility, visit ibidi.com/chemicals.

Chemical / Solvent	Compatibility
Methanol	Yes
Ethanol	Yes
Formaldehyde	Yes
Acetone	No
Mineral oil	Yes
Silicone oil	Yes
Immersion oil	See Section “Immersion Oil”

Immersion Oil

The compatibility with immersion oil depends on the used substrate.

For research use only!

Further information can be found at ibidi.com. For questions and suggestions, please contact us by e-mail at info@ibidi.com or by telephone at +49 (0)89/520 4617 0.